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# Phytochemical Investigation of *Dicoma tomentosa* to Isolate Secondary Metabolites

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ABSTRACT: *Dicoma tomentosa* Cass. (Family: Asteraceae) is an erect, annual herb of dry fields and sandy arid places. The whole plant is strongly bitter and is used for different ailments. Phytochemical investigation of the aerial part of plant was done to screen its secondary metabolites. Sesquiterpene lactones belonging to melampolide group of germacranolides were found to be characteristic of the species and were isolated along with triterpenes and steroids.

Keywords: Dicoma tomentosa; Natural products; Sesquiterpenoids; Spectral analysis.

**INTRODUCTION:** Several products isolated from plants, marine flora and microorganisms possess the unique properties like greater number of chiral centers, increased steric complexity, fewer hetero atoms, fewer heavy atoms etc. It accounts for the huge structural diversity of natural products which further results into its rich bioactive potentiality<sup>1</sup>. The genera Dicoma belonging to the family Compositae, tribe-Mutisieae and subtribe- Gochanatiinae<sup>2</sup> is of great medicinal and economic importance. For intangible, physiological and structural reasons, the group seems to be well adapted to many habitats such as deserts, which are difficult for other plants. As a result the Compositae includes innumerable successful species<sup>3</sup>.A large numbers of plants are ornamental and cultivated in gardens and homes. Some of them have medicinal importance. The genus Dicoma with approximately 35 species distributed over tropical South Africa with one species in Asia. Most of species are herbs or small shrubs but even trees are known. D. tomentosa Cass. is used medicinally in Western Africa; D. anomala Sond., D. capensis Less., D. speciosa E. May., D. zeyheri Sond are used in South Africa<sup>4</sup>. D. tomentosa is used as tooth cleaner<sup>5</sup>. Its leaves are used in the neighbourhood of Belgium as a febrifuge, during febrile attacks in the mother after child birth<sup>6</sup>. In Africa it is used as a local application to putrescent wounds. It is reported to possess prominent antiplasmodial activity<sup>7-9</sup>. The versatile features of secondary metabolites prompted us to study these phytochemicals from the plant D. tomentosa. The shed air dried aerial part of the plant was screened chemically to investigate its secondary metabolites.

## MATERIALS AND METHODS:

**General experimental procedures:** Active neutral (Acme Synthetic Chemicals) and silica gel (BDH, 60-120 mesh) were used as adsorbent for column chromatography. Qualitative and quantitative thin layer chromatographies were conducted on TLC aluminium sheets Kieselgel 60  $F_{254}$  (E. Merk). Preparative TLC performed on TLC glass plates silica gel 60  $F_{254}$  precoated (20×20 cm) layer thickness 0.5mm (E. Merk). Spots on TLC plates were visualized by spraying with 2% cerric ammonium sulphate in 2N  $H_2SO_4$ .

Melting points were determined in soft glass capillaries in an electrothermal melting point apparatus and are uncorrected. The identities of compounds were confirmed by comparison of their melting points and spectral data with literature values and also by mixed mp, co-TLC and co-IR with authentic samples.

The reported UV and IR spectra were recorded on Hitachi U-2000 double beam spectrophotometer and FT-IR spectrophototometer Magna IR-550 respectively. <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded on JEOL FX 90 Q, JEOL AL 300, Bruker DRX 300 and Bruker WM 400 instruments using CDCl<sub>3</sub>, C<sub>6</sub>D<sub>6</sub>, d<sub>6</sub> DMSO as solvents and TMS as an internal reference. EIMS, CIMS and FABMS data were generated on JEOL D-300, Shimadzu QP-5000 and JEOL-JMS-SX 102A spectrometers respectively.

Plant materials were collected from Sawai Madhopur and Jaipur regions of Rajasthan and it was characterized by the Herbarium In-charge, Department of Botany, University of Rajasthan, Jaipur.

Extraction and isolation of constituents from aerial parts of *Dicoma tomentosa*: Air dried and coarsely powdered plant material (3kg) was extracted with  $CH_2Cl_2$ -ether-mathanol (1:1:1) mixture in cold for 12 hrs. The extract was concentrated at reduced pressure

and subjected to column chromatography over silica gel. The eluants used and fractions obtained are cited in table 1 to isolate different secondary metabolites from the plant material (Figure 1).

Table 1:	Eluants and the fractions from column
	chromatography.

Fraction	Eluant	Residue left after re- moval of solvent		
No.		Nature	Amount	
1.	Petroleum-ether	Yellow Oil	150 mg	
2.	Petroleum- ether: Benzene (3:1)	Yellow semisolid	160 mg	
3.	Petroleum- ether: Benzene (1:1)	Yellow solid	500 mg	
4.	Benzene	Yellow green solid	150 mg	
5.	Benzene: EtOAc (3:1)	Yellow gum	200 mg	
6.	Benzene: EtOAc (1:1)	Yellow gum	190 mg	
7.	EtOAc	Yellow solid	120 mg	

**RESULTS AND DISCUSSION:** The air-dried and coarsely powdered aerial parts were extracted with dichloromethane-ether-methanol (1:1:1) mixture in cold. The resulting extract was concentrated at reduced pressure and chromatographed over silica gel. Subsequent fractions (Table 1) on preparative TLC gave following compounds.

Fraction 1 could not be investigated further on account of complex mixture of several non polar compounds as evidenced by the appearance of trailing on TLC plate. Fractions 2-7 were found to be a mixture of two or more compounds and each fraction was subjected to preparative TLC to get compounds in pure state.

**Component from fraction 2:** Fraction 2 showed a major spot along with some very minor spots on qualitative TLC plate. It was therefore purified by preparative TLC using petroleum-ether: benzene (2:1) as solvent system.

**1. Isolation of Lupeol acetate:** It was isolated as colourless crystals 110 mg, mp 218-19 <sup>0</sup>C and belongs to lupane series of triterpenes. It responded to positive Noller test for triterpenes.

IR v <sub>max.</sub> [KBr]	1725 (OAc) , 1640 (C=C) , 1160
cm <sup>-1</sup>	, 990 cm <sup>-1</sup> (C-O stretch).
<sup>1</sup> H NMR [300	1.69s (Me-C=C), 4.69 sbr and
MHz,	4.57 s (Vinylic protons), 4.38dd
CDCl <sub>3</sub> , δ ppm]	(J=12, 5HZ, H-3), 2.38m (H-19),

	2.10s (OAc), 0.76, 0.79, 0.83,
	0.94, 0.97 and 1.03 (all singlets
	representing six methyl groups).
MS (m/z)	468 $[M]^+$ (C <sub>32</sub> H <sub>52</sub> O <sub>2</sub> ), 453 [ M-
	Me ] $^{+}$ , 450 [ M-H <sub>2</sub> O] $^{+}$ , 408 [
	M-AcOH ] <sup>+</sup> , 242, 240, 231 and
	213, etc.

**Components from fraction 3:** Fraction 3 revealed the presence of two compounds on TLC plate. These compounds were separated by prep. TLC in petroleum-ether: benzene (1:1) mixture.

**2. Isolation of Stigmasterol:** It was the second component separated from fraction 3 and was obtained as shining needles, 175mg. mp 166-67  $^{0}$ C after crystallization from CHCl<sub>3</sub> – methanol (1:1) mixture. It gave positive Liebermann–Burchard and Salkowski test for sterol and TNM test for unsaturation.

IR $v_{max}$ . KBr	3400-3200 (OH), 1460 (-
	CH=CH- bending), 1380,
	1360, 1260, 1050, 960, 800
	$cm^{-1}$ .
<sup>1</sup> H NMR [300 MHz	5.34 tbr (H-6), 5.04 dd(J=16,
	10 Hz, H-22), 5.12 dd
	(J=16,10 Hz, H-23), 3.52 m
	(H-3α), 0.81 t (J=7 Hz, C-29
	methyl), 0.91 d (J=7 Hz, C-21
	methyl), 1.56 s (OH), 1.16 s
	(C-27 methyl), 0.88 s (C-19
	methyl), 0.79 s (C-18 methyl).
MS (m/z)	412 [M] <sup>+</sup> (C <sub>29</sub> H <sub>48</sub> O), 397 [M-
	Me] <sup>+</sup> , 328, 314, 302, 300, 271,
	253.

**3.** Isolation of  $\beta$ -Sitosterol: Yellowish solid obtained from fraction-3 after removal of solvent gave a colourless solid, 275 mg which was crystallized from benzene as colourless flaps, mp 136-37  $^{\circ}$ C and characterized as  $\beta$ -sitosterol.

#### **Components from fraction 4:**

**4. Isolation of Betulin:** The fraction 4 afforded yellowish solid after removal of solvent. After crystallization from CHCl<sub>3</sub>-methanol (1:1) betulin was isolated as colourless needles, 120 mg, mp 253-54  $^{\circ}$ C (lit 254-56  $^{\circ}$ C). It belongs to lupane series of triterpenes and gave positive Noller test. It also developed yellow colour with tetranitromethane indicating its unsaturated nature.

**Components from fraction 5:** The fraction 5 showed the presence of three compounds on TLC plate. Recolumn chromatography followed by prep. TLC of this fraction afforded following compounds **5. Isolation of Taraxasterol:** It was obtained from first fraction of re-column chromatography and was crystallized from methanol as white needles, 40 mg, mp 225-226 <sup>0</sup>C. It was highly soluble in benzene, chloroform and acetone. It responded positively to TNM test (yellow colour). It also gave pink-violet colour with Libermann-Burchard reagent and red colour with chlorosulphonic acid. It also responded positively to Salkowski and Noller tests. It displayed following spectral details:

IR v <sub>max</sub> [KBr]	3470 (OH), 2980, 2960,
	2855 (C-Hstretch), 1650
	(c=c stretch), 1460, 1375
	(gem dimethyl groups) and
	$1050 \text{ cm}^{-1}$ (c-o stretch).
<sup>1</sup> H NMR [300 MHz,	0.88 s, 0.87 s, 0.86 s, 0.85 s,
$CDCl_3, \delta(ppm)]$	0.84 s, 0.83 s, 0.82 d (J=7
	Hz) (these signals corre-
	sponded to seven methyl
	groups), 4.56 m
	(exomethylene protons),
	3.23 dd (J=7,11 Hz, H-3).
MS (m/z)	$426[M]^+$ (C <sub>30</sub> H <sub>50</sub> O), 411
	$[M-Me]^+$ , 408 $[M-H_2O]^+$ , 203
	etc.

6. Isolation of 8-Desoxyurospermal A -15-Oacetate. It was the second component isolated from fraction 5. It was obtained as colourless gum, 70 mg. It gave single spot on TLC plate. Its melting point could not be determined due to its sticky nature. It displayed following spectral details:

IR $v_{max}$ CHCl <sub>3</sub>	2950-2840, 1775 (γ-lactone),
cm <sup>-1</sup>	1740 (OAc), 1670, 1635 (C=C-
	CHO), 1470, 1390, 1320, 1240,
	1145, 1100, 1040, 980, 910.
<sup>1</sup> H NMR [300	6.52 ddd (J=7, 8.5, 1.5 Hz, H-1),
MHz,	2.49 m (H-2), 2.32 m (H-2'), 2.61
$CDCl_3, \delta(ppm)]$	ddd (J=6, 2, 12.5 Hz, H-3), 2.11
	brdd (J=12, 12.5Hz , H-3'), 5.29
	brd (J=10Hz, H-5), 4.67 dd (J=10,
	11Hz, H-6), 2.28 ddddd (J=3, 3.5,
	7, 10, 11Hz, H-7), 2.95 dddd
	(J=3, 10, 6.5, 12Hz, H-8), 1.52 m
	(H-8 <sup>'</sup> ), 2.43 brdd (J=10, 14Hz, H-
	9), 2.5 m (H-9'), 6.20d (J=1.5Hz,
	H-13), 5.50 d (J=1.5Hz, H-13'),
	9.48 s (H-14), 4.79 d (J=12.5 Hz,
	H-15), 4.71 d (J=12.5 Hz, H-15').
MS m/z (rel. int	$304 [M]^+ (0.1) (C_{17}H_{20}O_5),$
%)	244.110 [M-HOAc] <sup>+</sup> (100), 216
,	[244-CO] <sup>+</sup> (24), 215 [244-CHO] <sup>+</sup>
	(37).

7. Isolation of 8-Desoxyurospermal A: It was the third component isolated from fraction 5 after

recolumn chromatography. It was obtained as colourless gum, 80 mg. It belongs to melampolide group of sesquiterpene lactones. It exhibited following spectral data:

$IR  \nu_{max} CHCl_3  cm^{\text{-}1}$	3500 (OH), 2970-2850, 1770 (γ-lactone), 1690-1630 (C=CCHO), 1600, 1380, 1285, 1175, 1100, 1070, 940,
<sup>1</sup> H NMR [300MHz,	6.54 ddd (J=7, 9, 1.5 Hz, H-1), 2.49 m (H-2), 2.33 m (H-2), 2.76 ddd (J=6, 2, 12 Hz, H-3), 2.03 ddd (J=3,11,12 Hz, H-3), 5.18 brd (J=10 Hz, H-5), 4.76 dd (J=10, 11 Hz, H-6), 2.28 ddddd (J=3, 3.5, 6.5, 10, 11 Hz, H-7), 2.94 dddd (J=3, 10, 6.5, 12 Hz, H-8), 1.52 dddd (J=3.5, 10, 6.5, 12 Hz, H-8), 2.42 ddd (J=6.5, 10, 14 Hz, H- 9), 2.11 ddd (J=6.5, 10, 14 Hz, H- 9), 6.20 d (J=1.5 Hz, H- 13), 5.50 d (J=1.5Hz,H-13), 9.48 s (H-14), 4.47 d (J=12.5 Hz, H-15), 4.29 d (J=12.5 Hz, H-15).
MS m/z (rel.int.%)	262 $[M]^+$ (2) $(C_{15}H_{18}O_4)$ , 224.110 $[M-H_2O]^+$ (8) $(C_{15}H_{16}O_3)$ , 216 $[244-CO]^+$ (12), 215 $[244-CHO]^+$ (15), 53 (100).

**Components from fraction 6:** Fraction 6 of column chromatography on evaporation of solvent furnished a residue. It showed the presence of two components, close in their  $R_f$  values. It was therefore, subjected to prep. TLC (benzene:  $CH_2Cl_2$ : ether, 4 : 4 : 1) to get compounds in pure state.

**8. Isolation of Urospermal A-15-O-acetate:** The first band of prep. TLC afforded urospermal A-15-O-acetate as colourless solid, 60 mg. It belongs to melampolide group of sesquiterpene lactones. Its spectral analysis showed the following results.

IR $v_{max}$ CHCl <sub>3</sub> cm <sup>-1</sup>	3420 (OH), 2950-2840, 1770
	$(\gamma$ -lactone), 1750 (OAc),
	1690, 1630 (C=CCHO),
	1420, 1240, 1170 (C-H bend-
	ing), 1140, 1030, 980, 900
	$(C=CH_2 bending).$
<sup>1</sup> H NMR [300 MHz,	6.78 brdd (J=7, 9 Hz, H-1),
CDCl <sub>3</sub> , δ(ppm)]	2.55ddd (J=6, 2, 12
	Hz, H-3), 2.25 brdd (J=11,12
	Hz, H-3), 5.15 br d (J= 10)
	Hz, H-5), 4.51 dd (J=10, 11

	Hz, H-6), 2.49 dddd (J=3,		
	3.5, 10, 11 Hz, H-7), 3.91 ddd	<sup>1</sup> H NMR[300 MHz,	6.86 brdd (J=7, 9 Hz, H-
	(J=3, 10, 6.5 Hz, H-8), 2.51	CDCl <sub>3</sub> , δ(ppm)]	1), 2.63 m (H-2), 2.37 m
	m (H-9), 6.45 dd (J=1.5, 3.5		(H-2 <sup>'</sup> ), 2.65ddd (J=6, 2, 12
	Hz, H-13), 6.23dd (J=1.5, 3		Hz, H-3), 2.8 brdd (J=11,
	Hz, H-13'), 9.39 s (H-14),		12 Hz, H-3 <sup>'</sup> ), 5.09 br d (J=
	4.74 d (J=12.5 Hz, H-15),		10 Hz, H-5), 4.57 dd
	4.63 d (J=12.5, H-15'), 5.64 d		(J=10, 11 Hz, H-6), 2.45
	(J=11 Hz, OH), 2.10 s (three		dddd (J=3, 3.5, 10, 11 Hz,
	protons of acetate group).		H-7), 3.87 ddd (J=3, 10,
MS m/z (rel. int. %)	320 $[M]^+$ (0.5) $(C_{17}H_{20}O_6)$ ,		6.5 Hz, H-8), 2.49 m (H-
	260 $[M-HOAc]^+$ (55), 242		9), 6.52 dd (J=1.5, 3.5 Hz,
	$[260-H_2O]^+$ (25), 214 [242-		H-13), 6.33 dd (J=1.5, 3
	CO] <sup>+</sup> (31), 213 [242- CHO] <sup>+</sup>		Hz, H-13'), 9.45 s (H-14).
	(31), 69 (100).	MS (m/z)	278 $[M]^+$ (C <sub>15</sub> H <sub>18</sub> O <sub>5</sub> ), 260
			[M-H <sub>2</sub> O]+, 232 [260-

**9. Isolation of Urospermal A:** The second band of prep. TLC gave urospermal A as colourless crystalline solid, 80 mg, mp 163-165 <sup>0</sup>C. Spectroscopic studies showed the following result:

IR	$v_{max}$ CHCl <sub>3</sub> cm <sup>-1</sup>	3540	(OH),	2990	-2850,
		1770		(y-lac	ctone),
		1680,1	1635	(C=C-0	CHO),
		1645,	1440,	1280,	1150,
		1080,	960, 90	$0 \text{ cm}^{-1}$ .	

CO]<sup>+</sup>, 231 [260-CHO]<sup>+</sup>.

## **Components from fraction 7:**

**10. Isolation of Betulinic acid:** Fraction 7 revealed the presence of single major compound on TLC examination. It was obtained as colourless crystals, 90 mg, mp 314-316  $^{\circ}$ C. It answered positive Noller's test. It furnished following spectral data:



Figure 1: Isolated secondary metabolites from the aerial part of D. tomentosa.

IR $v_{max}$ KBr	3425-2800 (OH, COOH),
	$1715(C=O), 1640, 900 \text{ cm}^{-1}.$
<sup>1</sup> H NMR [90 MHz,	4.68 sbr and 4.56 sbr
$CDCl_3, \delta(ppm)]$	(=CH <sub>2</sub> ), 1.68 s (=C-CH <sub>3</sub> ),
	2.30 m (H-19), 3.27 dd
	(J=12, 5 Hz, H-3α), 0.76 s
	(3H), 0.78 s (3H), 0.82 s
	(3H), 0.96 s (3H), 1.03 s
	(3H) for five tertiary
	methyl groups.
MS (m/z, rel. int. %)	456 $[M]^+$ (C <sub>30</sub> H <sub>48</sub> O <sub>3</sub> ), 438
•	$[M-H_2O]^+$ (22), 423 [438-
	$Me]^+$ (14), 411 [M-
	COOH] <sup>+</sup> (12), 410 (9), 342
	$[M-C_6H_{10}O_2]^+$ (7.2), 248
	(65), 220 (42), 207 (79),
	203 (49), 189 (100), 149,
	69, etc.
	·

**CONCLUSION:** The aerial part of *Dicoma tomentosa* was phytochemically screened to observe different natural products of the plant. Four highly oxygenated germacranolides, three triterpenes (among them two of lupane series), two steroids were isolated and characterized with the help of spectral studies.

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## **REFERENCES:**

1. Sharma V., Sharma T., Kaul S., Kapoor K. K., Dhar M. K. (2017) Anticancer potential of labdane diterpenoid lactone "andrographolide" and its derivatives: a semi-synthetic approach, *Phytochemistry Reviews*, 16(3), 513-26.

- 2. Goyal P. K., Aggarwal R. R. (2013) A Review on Phytochemical and Biological Investigation of Plant Genus *Pluchea*, *Indo American Journal of Pharm Research*, 3(4), 3000-7.
- **3.** Stepp J. R., Moerman D. E. (2001) The importance of weeds in ethnopharmacology, *Journal of Ethnopharmacology*, 75(1), 19-23.
- 4. Abdillahi H. S., Van Staden J. (2013) Application of medicinal plants in maternal healthcare and infertility: a South African perspective, *Planta medica*, 79(07), 591-9.
- 5. Asolkar L. V., Kakkar K. K., Chakre O. J. (1956) Glossary of Indian Medicinal Plants with active principal, part-I. Publications and Information Directorate, Council of Scientific and Industrial Research Publ, New Delhi, 165.
- 6. Jain S. K. (1994) Ethnobotany and research in medicinal plants in India, *Ethnobot. Search, New Drugs*, 1,185,153-68.
- Jansen O., Tits M., Angenot L., Nicolas J. P., De Mol P., Nikiema J. B., Frédérich M. (2012) Antiplasmodial activity of *Dicoma tomentosa* (Asteraceae) and identification of urospermal A-15-O-acetate as the main active compound, *Malaria journal*, 11(1),289.
- 8. Abdillah S., Tambunan R. M. Farida, Y. Sandhiutami, N. M. D. Dewi, R. M. (2015) Phytochemical screening and antimalarial activity of some plants traditionally used in Indonesia, *Asian Pacific Journal of Tropical Disease*, 5(6), 454-457.
- **9.** Kaur R., Kaur, H. (2017) Plant Derived Antimalarial Agents. *Journal of Medicinal Plants*, 5(1), 346-363.