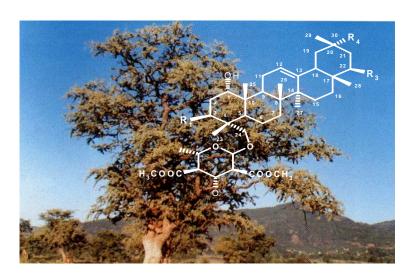
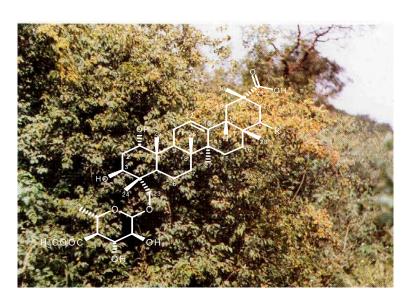
# Isolation and characterization of anti-bacterial compounds present in members of *Combretum* section, Hypocrateropsis



Combretum imberbe



Combretum padoides

08-11-05

J.E Angeh

# Isolation and characterization of antibacterial compounds present in members of *Combretum* section, Hypocrateropsis

J.E Angeh

B.Tech, MSc (Bauchi)

Submitted in fulfilment of the requirements for the degree of Doctor of Philosophy (PhD)

in the

# Phytomedicine Programme, Department of Paraclinical Sciences Faculty of Veterinary Science



Promoter: Prof. J.N. Eloff

Co-Promoter: Prof. G.E. Swan

Date of submission: November 2005

# **DECLARATION**

The experimental work described in this thesis was conducted in the Phytomedicine Programme, Department of Paraclinical Science, Faculty of Veterinary Science, University of Pretoria between June 2002 to April 2005 and at the Department of Molecular Natural Products Research, Hans-Knöll Institut fur Naturstoff Forschung (Hans-Knöll Institute for Natural Product Research) Jena, Germany from March 2003 to June 2003, under the supervision of Prof. JN Eloff, Prof. G. Swan and Dr. Isabel Sattler.

These studies are the result of my own investigations, except where the work of others is acknowledged, and has not been submitted in any other form to another University. I declare the above statement to be true.

JE Angeh

#### **ACKNOWLEDGEMENTS**

I would like to express special thanks to my supervisors, Prof. JN Eloff and Prof. GE Swan for supervising this research, and for their invaluable advice and encouragement during the time we have worked together. Many thanks to the Hans-Knoll Institute (HKI) Jena, Germany for determining some of the assays (cytotoxicity, anti-inflammatory and antiviral assays) that were needed for this research. The research staff of (HKI), especially members of the Molecular Natural Product Research group are gratefully acknowledged for their time and expertise that was always generously given during my research stay in HKI from March 2004 to June 2004. Special appreciation is due to my friends Dr. S. Huang of HKI, Dr. David Katerere and Dr. Lyndy McGraw for their generous support during my research period. My greatest thanks go to the National Research Foundation (NRF), Germany Academic Exchange Service (DAAD), Phytomedicine Programme and the Faculty of Veterinary Science, University of Pretoria for financial support during these studies.

Special thanks to my supervisor's wife (Mrs Christna Eloff) for her motherly support to my family during this period and my most profound appreciation goes to my loving wife Irene, my daughter Sandy and my late father (Angeh David Ekwa) and mother (Lydia Anchi Angeh) who had been a springboard in my academic support.

#### LIST OF ABBREVIATIONS USED

1D 1-dimentional 2D 2-dimentional

CC Cytotoxic concentration
COSY Correlation spectroscopy

DCM Dichloromethane

DEPT Distortionless enhancement by polarization transfer

DMSO Dimethyl sulfoxide
El Electron impact

ESI Electron spray impact
GI Growth inhibition

HMBC Heteronuclear multiple bond correlation
HMQC Heteronuclear multiple quantum coherence

IC Inhibition concentration

IUPF Indigenous Plant use Forum

MIC Minimum inhibitory concentration

MS Mass spectrometer

NMR Nuclear magnetic resonance

NOESY Nuclear overhayser enhancement spectroscopy

n-phase normal phase

NCCLS National committee for clinical laboratory standards

TDH Threonine dehydrogenase

B1 Bacillus subtilis ATTC 6633 (IMET) NA
B2 Bacillus subtilis ATTC 6633(IMET) AS

B3 Staphylococcus aureus (IMET 10760) SG 511

B4 Escherichia coli SG 458

B9 Pseudomonas aeruginosa K799/61

M2 *Mycobacterium smegmatis* SG 987 (HK10056)

M4 *Mycobacterium vaccae* IMET 10670

H4 Sporobolomyces salmonicolor SBUG 549

H8 Candida albicans BMSY 212
P1 Penicillium notatum JP 36

HKI Hans-Knoll Institute

VLC Vacuum liquid chromatography

TLC Thin layer chromatography

HPLC High performance liquid chromatography
EMW Ethyl acetate, methanol, water (40:5.4:4)
BEA Benzene, ethyl acetate, ammonia (90:10:1)
CEF Chloroform, ethyl acetate, formic acid (5:4:1)

INT p-iodonitrotetrazolium violet

C.I Combretum imberbe
C.P Combretum padoides

C.Cs.C Combretum celastroides ssp. celastroides
C.Cs.O Combretum celastroides ssp. orientale

TDH Threonine dehydrogenase

EtOAc Ethyl acetate

Retention factor

SA Staphylococcus aureus
EF Enterococcus faecalis

EC Echerichia coli

PA Pseudomonas aeruginosa

ssp Subspecies

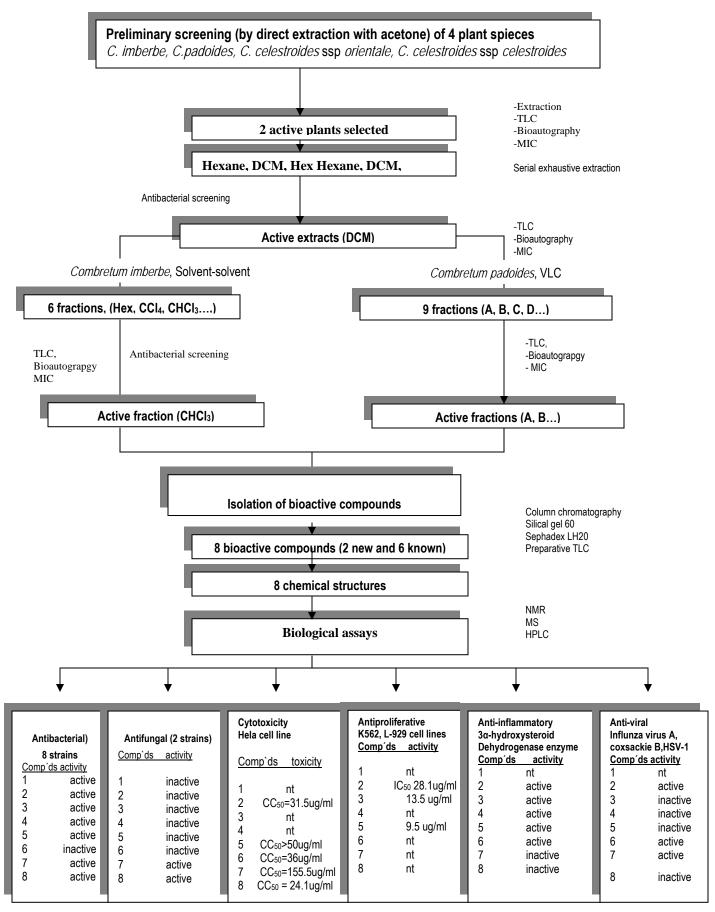
#### **ABSTRACT**

Combretum imberbe (leadwood, hardekool) has been used for medicinal purpose and several studies have been carried out to investigate the chemical compounds present in the bark of this plant. Preliminary experiments indicated that the leaves of this plant contain antibacterial compounds that do not occur in other Combretum species. Leaves of C. imberbe and the closely related C. padoides belonging to the Combretum section, Hypocrateropsis of the African Combretaceae were extracted and fractionated by bioassay-guided fractionation yielding 5 compounds.

Compounds	$R_1$	$\mathbf{R}_2$	$\mathbf{R}_3$	$R_4$
1	OH	Н	Н	СООН
2	H	Η	Η	COOH
3	H	Н	=0	CH <sub>2</sub> OH
4	H	Н	OH	COOH
5	2,4-Di-Ac-O-Rh	OH	Η	Н

Two new antibacterial pentacyclic triterpenoids ( $1\alpha$ ,  $24\beta$ -dihydroxyl-12-oleanen-29-oic acid- $3\beta$ -O- $\alpha$ -2, 4-diacetylramnopyranoside and  $1\alpha$ ,  $3\beta$ -dihydroxyl-12-oleanen-29-oic acid- $23\beta$ -O- $\alpha$ -4-acetylramnopyranoside) **5** and **6** along with six known triterpenoids **1-4** ( $1\alpha$ ,  $3\beta$ -dihydroxyoleanen-12-29-oic, 3-hydroxyl-12-olean-30-oic, 3, 30-dihydroxyl-12-oleanen-22-one, 1,3, 24-trihydroxyl-12-olean-29-oic acid 7 ( $1\alpha$ ,  $22\beta$ -dihydroxyl-12-oleanen-30-oic acid) and **8** (24-ethylcholesta-7, 22,25-trien-3-ol-O- $\beta$ -D-glucopyranoside) were isolated with the aid of closed and open column chromatography.

Compound	R¹	R <sup>2</sup>
6	4-Ac-O-Rh	Н
7	Н	OH



All eight compounds had moderate (MIC of 60  $\mu$ g/ml) to strong (10  $\mu$ g/ml) antibacterial activity against *Staphylococcus aureus, Bacillus subtilis, and Mycobacterium vaccae* with **2, 5** and **7** being most active. Compound **2** and **3** also had strong anti-inflammatory activity against 3 $\alpha$ -hydroxylsteriod dehydrogenase enzyme with an IC  $_{50}$  of 10  $\mu$ g/ml and 7.8  $\mu$ g/ml as well as moderate cytotoxicity (CC $_{50}$  = 17.6  $\mu$ g/ml and CC $_{50}$  = 10.5  $\mu$ g/ml) against Hella cell lines. Compound **2** and **5** also had moderate anti-proliferative (GI $_{50}$  =16.5  $\mu$ g/ml, 13.2  $\mu$ g/ml, 8.7  $\mu$ g/ml) activity against K-562, L-929 cell lines.

The structures of the compounds were elucidated on the basis of 1-dimensional (1D) and 2-dimesional (2D) Nuclear Magnetic Resonance (NMR) experiments, as well as Electron Impact (EI) and Electron Spray Impact (ESI) mass spectrometric techniques.

The results of this study have added new compounds to the global database of phytocompounds, expanded our knowledge on the phytochemistry of *Combretum*, confirmed the rationale of the ethnomedicinal use of *C. imberbe*, and opened up potential new applications of extracts.

#### PAPERS PREPARED FROM THIS THESIS

Angeh, J. E., Eloff, J. N., Swan, G.E., 2005. Comparing two extraction methods in isolating antibacterial compounds from *Combretum* section Hypocrateropsis (Manuscript).

Angeh, J. E., Eloff, J. N., Swan, G.E., Huang S., Sattler I., 2005. Activity guided isolation of a new anti-microbial triterpenoid from Combretum imberbe. (Manuscript).

Angeh, J. E., Eloff, J. N., Swan, G.E., Huang S., Sattler I., 2005. Novel anti-microbial triterpenoid from *Combretum padoides* (Manuscript).

Angeh, J. E., Eloff, J. N., 2005. Synergism in isolated natural compounds (Manuscript).

#### **CONFERENCES AND PROCEEDINGS**

#### 2003

#### Indigenous Plant Forum (IPUF), Kloof Avenue Rustenburg (South Africa)

Poster: JE Angeh, GE Swan, JN Eloff. Comparing two extraction methods in isolating antibacterial compounds from *Combretum Section* Hypocrateropsis.

#### Faculty day, Faculty of Veterinary Science, University of Pretoria.

Poster: JE Angeh, GE Swan, JN Eloff. The significance of serial exhaustive extraction in isolating antibacterial compounds from *Combretum imberbe*.

#### 2004

#### Indigenous Plant Forum (IPUF), Clanwilliam (South Africa).

Paper: JE Angeh, GE Swan, JN Eloff, S. Huang, I. Sattler. Bioassay-guided isolation of pentacyclic triterpenoids from *Combretum imberbe*.

#### Faculty day, Faculty of Veterinary Science, University of Pretoria.

Paper: JE Angeh, GE Swan, JN Eloff, S. Huang, I. Sattler. Novel biologically active triterpenes from Combretum imberbe and Combretum padoides

#### 2005

# Annual conference of Society for Medicinal Plants Research, Florence, Italy

Paper: J. Angeh, J. Eloff, G. Swan, S. Huangi I, Sattler. Novel biologically active triterpenoids from the African Combretaceae

# Indigenous Plant Use Forum (IPUF), Gramhamstown

Paper: J. Angeh, J. Eloff. Synegetic effect of triterpenoids isolated from *Combretum* species.

# **TABLE OF CONTENT**

DECLARATION		i
ACKNOWLEDGEMEN	ITS	i
LIST OF ABBREVIATION	ONS USED	iii
ABSTRACT		iv
PAPERS PREPARED	FROM THIS THESIS	ν
CONFERENCES AND	PROCEEDINGS	vi
CHAPTER 1. INTROD	UCTION	1
1.1 Introduction		1
1.2 Hypothesis		2
1.3 Aim of research		3
1.3.1 Objectives		3
CHAPTER 2: LITERA	TURE REVIEW	4
2.1 Introduction		4
2.2 Antibiotic resistanc	e	5
2.3 Natural products in	drug discovery	6
2.4 Plants as a potentia	al source of antibiotics	9
2.5 Plants and antibact	terial production	11
2.6 The familly Combre	etaceae	13
2.6.1 Taxonomy		13
2.6.2 Taxonomy of	the section Hypocrateropsis	14
2.6.2.1 <i>Com</i>	nbretum imberbe Engl. & Diels	14
2.6.2.2 C <i>om</i>	nbretum padoides Engl. & Diel	15
2.6.2.3 <i>Con</i>	mbretum celastroides Welw Ex M.A. Lawson	16
2.7 Ethnopharmacolog	y of Combreteceae	17
2.8 Phytochemistry/che	emistry and biological activity of Combretaceae	18
2.8.1 Tannins		19
2.8.1.1 Clas	sses of tannins	19
28111 Hvdr	rolysable tannins (HTs)	19

2.8.1.1.	2 Pr	roanthocyanidins (Condensed tannins)	20
2.8.2	Tannins ch	emistry of Combreteceae	22
2.8.3	Stilbenoids		22
2.8.4	Flavonoids		25
2.8.4.1.	2 FI	avonoids chemistry of Combretaceae	26
2.8.5	Terpenoids		27
2.8.5.1	CI	asses of terpenes	28
2.8.5.1.	1 Te	erpene essential oils	28
2.8.5.1.	2 Di	terpenoids and Gibberellins	28
2.8.5.1.	3 Tr	iterpenoids and Steroids	29
2.8.5.1.	4 Te	etraterpenoids (Carotenoids)	29
2.8.5.2	Te	erpenoids chemistry of Combretaceae	30
2.8.6	Alkaloids		32
2.8.6.1	CI	asses of alkaloids	33
2.8.6.1.	1 O	rnithine and lysine derivatives	33
2.8.6.1.	2 Pł	nenylalanine and tyrosine derivatives	33
2.8.6.1.	3 Tr	yptophan derivatives	34
2.8.6.2	Co	ombretaceae alkaloids chemistry	34
2.8.7	Other cons	tituents	34
2.9	Methods de	eveloped and results obtained in the phytomedicine programme	35
2.9.1	Bioautogra	phy and MIC methods	35
2.9.2	Overview o	f activities in Combretaceae research in the phytomedicine group	35
2.9.2.1	Se	election of plants	35
2.9.2.2	Se	election of best extraction procedure	36
2.9.2.3	Se	election of best purification procedures	36
2.9.2.4	De	eveloping a novel way of determining antibacterial activity	36
2.9.2.5	Ar	ntibacterial activity of <i>C. erythrophyll</i> um	36
2.9.2.6	Ar	ntibacterial activity and stability of 27 members of Combretaceae	36
2.9.2.7	St	ability of antibacterial activity in C. erythrophy llum	36
2.9.2.8	Α	proposal for expressing antibacterial activity	37
2.9.2.9	) Iso	olation of antibacterial compound from C. woodii	37
2.9.2.10	) Is	olation of antibacterial compounds from C. erythrophyllum	37
2.9.2.11	O	ther biological activities of <i>Combretum</i> species	37

2.9.3 O	Other work done on <i>C. imberbe</i> and <i>C. padoides</i>	38
СНАРТ	TER 3: PLANT COLLECTIONS, EXTRACTION AND ANALYSIS	44
3.1 Intro	oduction	39
3.2 Mat	terial and Methods	39
3.2.1	Experimental design for bioassays	39
3.2.2	Plant collection.	40
3.2.3	Plant preparation and extraction	40
3.2.3.1	Direct extraction with acetone	40
3.2.3.2	Serial exhaustive extraction	41
3.2.4	Analysis of plant extracts for preliminary screening	42
3.3 Res	sults	43
3.3.1	Extraction	43
3.3.1.1	Direct extraction with acetone	43
3.3.1.2	Serial exhaustive extraction	43
3.3.2	TLC analysis of plant extracts for preliminary screening	44
3.3.3	TLC analysis of plant extracts from serial exhaustive extraction	45
3.4 Dis	cussion and conclusion	48
3.5 Sur	mmary	49
СНАРТ	TER 4: BIOLOGICAL ASSAYS FOR PRELIMINARY SCREENING	50
4.1 Intro	oduction	50
4.2 Mat	terials and methods	50
4.2.1	Antibacterial assay	50
4.2.1.1	Test organisms	50
4.2.2	Bioautography for preliminary screening	50
4.2.3	.Microplate dilution assay for preliminary screening	51
4.2.4	Determination of total activity	52
4.3 Res	sults	53
4.3.1	Antibacterial assays	54
4.3.1.1	Bioautography	57
4.3.1.2	Bioautography of serial exhaustive extraction extracts	57

4.3.2	Minimum inhibitory concentration of extracts from serial exhaustive extraction	68	
4.3.3	Total activity of extracts	60	
4.4 Discussion and conclusion			
4.5 Sur	nmary	61	
СНАРТ	TER 5: PRELIMINARY SEPARATION AND ISOLATION OF BIOACTIVE COMPOUNDS	63	
5.1 Intro	oduction	63	
5.2 Mat	erials and methods	63	
4.3.4	Solvent-solvent fractionation	63	
5.2.2	Column chromatography	63	
5.2.2.1	Vacuum Liquid chromatography	65	
5.2.2.2	Gravity gradient column chromatography	66	
5.2.2.3	Sephadex LH-20.	67	
5.2.3	Conventional Preparative TLC	68	
5.2.4	Analysis and Grouping of fractions	68	
5.2.4.1	Combination of fractions	69	
5.2.5	Dereplication	69	
5.3 Res	sults and discussions	69	
5.3.1	Solvent-solvent fractionation of the DCM extracts	69	
5.3.1.1	Complexity of fractions	70	
5.3.2	Vacuum liquid chromatography	71	
5.3.2.1	Fractions from VLC process of <i>C. imberbe</i>	74	
5.3.3	Gravity gradient chromatography fractions from solvent-solvent fractionation process of C. in	nberbe	
		74	
5.3.4	Sephadex LH-20	75	
5.3.5	Conventional preparative TLC	76	
5.3.6	Gravity gradient chromatography	76	
5.3.7	Bioautography and MIC	78	
5.3.8	Bioautography and MIC of VLC fractions	81	
5.3.9	9 Dereplication of compounds		
5.4 Sur	nmary	83	

CHAP	TER 6: INSTRUMENTAL ANALYSIS AND STRUCTURAL ELUCIDATION OF ISOLATED	
	COMPOUNDS	84
	oduction	
	clear magnetic resonance spectroscopy [NMR ]	
6.2.1	NMR sample preparation	84
6.2.2	One-dimensional spectroscopy	85
6.2.2.1	<sup>1</sup> H-NMR	85
6.2.2.2	<sup>13</sup> C-NMR	86
6.2.2.3	DEPT	86
6.2.3	Two-dimensional spectroscopy	86
6.2.3.1	Homonuclear correlations	86
6.2.3.1	.1 <sup>1</sup> H- <sup>1</sup> H Correlation Spectroscopy (COSY)	86
6.2.3.2	Heteronuclear correlations	86
6.2.3.2	.1 Heteronuclear Multiple Quantum Coherence (HMQC) (¹H-¹³C COSY)	87
6.2.3.2	.2 Heteronuclear Multiple Bond Connectivity (HMBC) (Long range <sup>13</sup> C- <sup>1</sup> H COSY)	87
6.2.3.2	.3 NOESY spectroscopy	87
6.3 Ma	ss spectrometry	88
6.3.1	Sample preparation	88
6.3.2	Gas Chromatography - Mass Spectrometry (GC-MS)	88
6.3.2.1	Electron Impact Spray MS (EIS-MS)	88
6.4 IR.		88
6.5 Res	sults and Discussions	89
6.5.1	Identification of compounds	89
6.5.1.1	Compound 1	89
6.5.1.2	Compound 2	94
6.5.1.3	Compound 3	95
6.5.1.4	Compound 4	97
6.5.1.5	Compound 5	99
6.5.1.6	Compound 6	104
6.5.1.7	Compound 7	110
6.5.1.8	Compound 8	114
6.6 Sur	mmary	119

CHAP	TER 7: BIOLOGICAL CHARACTERIZATION OF ISOLATED COMPOUNDS	121
7.1 Intr	oduction	121
7.2 Ma	terial and methods	121
7.2.1	Anti-microbial activity	121
7.2.1.1	Antibacterial activity (Microplate dilution assay)	121
7.2.1.2	Antibacterial and antifungal activity (Agar diffusion assay)	121
7.2.1.3	Antimicrobial effect of some combined compounds	122
7.2.2	Anti-inflammatory activity	122
7.2.2.1	Preparation of Cytosol	123
7.2.2.2	Preparation of Purified $3\alpha$ -Hydroxysteroid Dehydrogenase	123
7.2.2.3	Enzyme Assays	123
7.2.2.4	Inhibition Studies	124
7.2.3	Antiproliferative and cytotoxicity assay	124
7.3 Res	sults and Discussion	125
7.3.1	Antibacterial activity (Microplate dilution assay)	125
7.3.2	Antimicrobial activity (Agar diffusion assay)	126
7.3.3	Combined antimicrobial effect of mixtures of isolated compounds	128
7.3.4	Anti-inflammatory activity	130
7.3.5	Anti-proliferative effect and Cytotoxicity	131
7.3.5	Structure activity relationship of isolated compounds	132
7.4 Sur	mmary	132
CHAP	TER 8: GENERAL CONCLUSION	134
8.1Intro	oduction	134
8.2 Eva	aluation on the best preliminary fractionation procedure	134
8.3 Iso	lation and chemical characterization of antibacterial compounds	135
8.4 Bio	logical characterization of plant species and isolated compounds	136
8.5 Eva	aluation of how well phytochemistry agrees with taxonomy based on anatomy	137
СНФБ	TER 0. REFERENCES	138-146

# **LIST OF FIGURES**

Figure 2-1: The sub generic classification for South Africa Combretaceae according to Carr (1988)
14
Figure 2-2: <i>C. imberbe</i> Leadwood/hardekool (Steyn, 1994)
Figure 2-3: <i>C. padoides</i> (Steyn, 1997)
Figure 2-4: <i>C. celastroides</i> ssp. <i>orientale</i> (Steyn, 1994)
Figure 3-1: Percentage of material extracted from <i>C. imberbe</i> and <i>C. padoides</i> by each solvent in the serial
exhaustive extraction process
Figure 3-2: Percentage of material extracted from <i>C. imberbe</i> and <i>C. padoides</i> by each solvent in the serial exhaustive extraction process
Figure 3-3: TLC chromatograms (viewed under UV 365 nm and vanillin sulphuric acid spay reagent) of
extracts from direct extraction with acetone in preliminary screening process
Figure 3-4: TLC chromatogram (viewed after spraying with vanillin sulphuric acid spray reagent) of seria
exhaustive extraction extracts of C. <i>imberbe</i> and <i>C. padoides</i> developed with EMW. Areas on the chromatogram cycled indicate UV active compounds
Figure 3-5: TLC chromatogram (viewed after spraying with vanillin sulphuric acid spray reagent) of seria
exhaustive extraction extracts of C. imberbe and C. padoides developed with CEF. Areas on the
chromatogram cycled indicate UV active compounds47
Figure 3-6: TLC chromatogram (viewed after spraying with vanillin sulphuric acid spray reagent) of seria
exhaustive extraction extracts of C. imberbe and C. padoides developed with BEA. Areas on the
chromatogram cycled indicate UV active compounds47
Figure 4-1: INT, coupling reagent for the colorimetric assay (reaction pathway for the assay of TDH)51
Figure 4-2: Bioautograms of acetone extracts of C. imberbe, C. padoides, C. celastroides ssp. orientale and
C. celastroides ssp. celastroides. TLC developed in EMW and sprayed with actively growing S
aureus cultures and later sprayed with INT. White areas indicate zones of growth inhibition 54
Figure 4-3: Bioautograms of acetone extracts of C. imberbe, C. padoides, C. celastroides ssp. orientale and
C. celastroides ssp. celastroides. TLC developed in EMW and sprayed with actively growing P
aeruginosa cultures and later sprayed with INT. White areas indicate zones of growth
inhibition54

Figure 4-4: Bioautograms of acetone extracts of <i>C. imberbe</i> , <i>C. padoides C. celastroides</i> ssp. orientale	and
C. celastroides ssp. celastroides. TLC developed in BEA and sprayed with actively growing S. au.	reus
cultures and later sprayed with INT. White areas indicate zones of growth inhibition	55
Figure 4-5: Bioautograms of acetone extracts of <i>C. imberbe</i> , <i>C. padoides</i> , <i>C. celastroides</i> ssp. orientale	and
C. celastroides ssp. celastroides. TLC developed in BEA and sprayed with actively growing E.	coli
cultures and later sprayed with INT. White areas indicate zones of growth inhibition	. 56
Figure 4-6: Bioautography of serial exhaustive extraction extracts on S. aureus sprayed with INT spra	ayed
reagent. White areas indicate zones of inhibition	57
Figure 4-7: Bioautography (BEA solvent system) of serial exhaustive extraction extracts against S. au	reus
sprayed with INT spray reagent. White areas indicate zones of inhibition	57
Figure 4-8: Bioautography of serial exhaustive extraction extracts against E. coli, using EMW solvent sys	stem
and sprayed with INT spray reagent. White areas idicate zones of inhibition	58
Figure 4-9: Bioautography of serial exhaustive extraction extracts against E.coli, using BEA solvent sys	stem
and sprayed with INT spray reagent. White areas indicate zones of ihibition	58
Figure 4-10: MIC of the hexane and DCM extracts of C. imberbe and C. padoides on S. aureus on microtitr	îе
plates. White wells indicate inhibition and purple wells indicate bacterial growth	58
Figure 4-11: MIC of the hexane and DCM extracts of <i>C. imberbe</i> and <i>C. padoides</i> against <i>E. coli</i>	58
Figure 5-1: Diagrammatic summary of the isolation routes	62
Figure 5-2: The protocol used for the solvent-solvent fractionation of the components in the DCM extract of	of C.
imberbe and C. padoides	64
Figure 5-3: Vacuum liquid chromatography apparatus (Houghton, 1998)	65
Figure 5-4: Separation of components present in the different fractions obtained by solvent-solvent extractions	ction
by EMW and sprayed with vanillin-sulphuric acid. Lanes from left to right: hexane, water, n-but	ane,
carbon tetrachloride, chloroform and 35% water in methanol fractions. In each case 50 $\mu \text{g}$ of 5 m $^{\circ}$	g/ml
stock solution was chromatographed.	71
Figure 5-6: Chromatogram ran in CEF indicating number of test tubes collected in the VLC process	71
Figure 5-7: Grouped fractions resulting from Vacuum Liquid Chromatography of DCM extract of C. imberbe	? ran
with CEF solvent system. Lane from left to right: DCM crude extract, fractions A, B, C, D, E, F	
G	72
Figure 5-8: Chromatogram from the combined fractions E, F and G ran on CEF solvent system	72
Figure 5-9: Quantities obtained from 6.55 g of chloroform fraction with the solvent-solvent fractional	ation
process of the DCM extract of C. imberbe	73

Figure 5-10: Chromatogram of fractions resulting from the wash of crystallized test tubes of fraction A and B run along the crude DCM extract
Figure 5-11: Separation of components present in 50 $\mu$ g of 12 different fractions resulting from 3 x 35 cm silications
gel 60 column using EMW as eluent and vanillin-sulphuric acid spray reagent
Figure 5-12: Quantities obtained from 6.55 g of chloroform fraction with the solvent-solvent fractionation
process of the DCM extract of <i>C. imberbe</i>
process of the Down extract of C. Imberbe
Figure 5-13: Chromatogram of the separation of 2.5 g of F <sub>8</sub> -F <sub>12</sub> combined fraction on Sephadex LH-20 ran with
chloroform/methanol (9:1) solvent system and spayed with vanillin sulphuric acid spray reagent
Figure 5-14: Compounds 4 and 5 isolated from <i>C. imberbe</i> by Sephadex LH-20 column
Figure 5-15: Gravity based separation of components of the chloroform fraction resulting from the solvent solvent fractionation of the DCM fraction of <i>C. padoides</i> ran in EMW and sprayed with vanilling sulphuric acid
Figure 5-16: Separation of re-grouped fractions of the gravity based separation of components of the chloroform fraction resulting from the solvent-solvent fractionation of the DCM fraction of <i>C padoides</i>
Figure 5-17: Chromatogram (developed in EMW sprayed with vanillin sulphuric acid spray reagent) o compounds isolated from <i>C. padoides</i>
Figure 5-18: Bioautogram of DCM extracts of <i>C. imberbe</i> (left) and <i>C. padoides</i> (right) leaves separated into different fractions by solvent-solvent extraction. TLC plate developed in EMW and sprayed with <i>S. aureus</i> culture incubated overnight and then sprayed with INT. Growth inhibition is indicated by colourless zones on TLC plates. Lanes from left to right: hexane, water, n-butanol, carbor tetrachloride, 35% water in methanol, chloroform fractions.
Figure 5-19: Bioautogram of DCM extracts of <i>C. imberbe</i> (left) and <i>C. padoides</i> (right) leaves separated into different fractions by solvent-solvent extraction. TLC plate developed in BEA and sprayed with <i>S. aureus</i> culture incubated overnight and then sprayed with INT. Growth inhibition indicated by colourless zones on TLC plates. Lanes from left to right: hexane, water, n-butanol, carbor tetrachloride, 35% water in methanol, chloroform fractions.
Figure 5-20: Bioautogram of the group separation (silica gel 60) of 6.55 g of the chloroform resulting from the
solvent-solvent separation of DCM extract of <i>C. imberbe</i> on EMW80
Figure 5-21: Bioautogram of the DCM extract of C. imberbe separated into different fractions in a Vacuum
Liquid Chromatography process. TLC plate developed in CEF (as best separating system) and
sprayed with S. aureus culture, incubated overnight and then sprayed with INT. Growth inhibition

indicated by colourless zones on TLC plates. Lanes from left to right: fractions A, B, C, D, E, F a	
Figure 5-22: Chromatogram of the crude extract of <i>C. imberbe</i> compaired with isolated compound dereplicate isolated compounds from the crude extract. Lanes from left to right: Crude extract	ls to
imberbe, compunds 1, 4, 5 and 3.	82
Figure 5-23: Identification of isolated compounds from the crude extract of <i>C. padoides</i>	
Figure 6-1: Skeletal unit of olean-12-ene type of pentacyclic triterpenoid isolated from <i>C. imberbe</i>	89
Figure 6-2: Fragmentation pattern of compound 1	91
Figure 6-3: EI-MS spectrum of compound 1	91
Figure 6-4: <sup>1</sup> H NMR spectrum of Compound1	92
Figure 6-5: <sup>13</sup> C NMR spectrum of Compound 1	93
Figure 6-6: <sup>13</sup> C NMR spectrum of Compound 2	95
Figure 6-7: Fragmentation patten of compound 3	96
Figure 6-8: Electron impact mass spectrum of Compound 3 indicating the m/e of each fragment	97
Figure 6-9: <sup>1</sup> H NMR spectrum of Compound 3	97
Figure 6-10: <sup>1</sup> H NMR spectrum of Compound 4	98
Figure 6-11: DEPT NMR spectrum of Compound 4 differentiating CH <sub>3</sub> -, CH <sub>2</sub> -, and CH- carbons	99
Figure 6-12: <sup>1</sup> H NMR spectrum of compound 5	102
Figure 6-13: DEPT spectrum of Compound 5 indicating the presence of CH <sub>3</sub> -, CH <sub>2</sub> - and CH- carbons in	in the
compound	103
Figure 6-14: <sup>1</sup> H NMR spectrum of compound 6	107
Figure 6-15: <sup>13</sup> C NMR of Compound 6	108
Figure 6-16: HMBC spectrum of Compound 6	109
Figure 6-17: <sup>1</sup> H NMR of compound 7	111
Figure 6-18: DEPT spectrum of Compound	112
Figure 6-19: <sup>13</sup> C NMR of Compound 7	113
Figure 6-20: <sup>1</sup> H-NMR of Compound 8	115
Figure 6-21: DEPT spectrum of Compound	
Figure 6-22: Compounds 1-5 isolated from <i>C. imberbe</i>	119
Figure 6-23: Compounds 6-8 isolated from <i>C. padoides</i> .	120

# **LIST OF TABLES**

Table 2-1: Plant-derived drugs widely employed in Western medicine (Adapted 1	rom Farnsworth
1984)	4
Table 2-2: Plants containing chemotherapeutic activity	7
Table 2-3: Some plant-derived preparations for medicinal use	10
Table 2-4: Derivatives of stilbenes (Combrestatins) isolated from Combretaceae	23
Table 2-5: Derivatives of dihydrostilbenes (Combrestatins) isolated from Combretaceae	23
Table 2-6: Derivatives of phenanthrenes isolated from Combretaceae	24
Table 3-1: Quantity extracted at each batch (A, B, C, D, E, and F) in a serial exhaustive extracted of <i>C. imberbe</i> leaves	_
Table 3-2: Quantity extracted at each batch (A, B, C, D, E and F) in a serial exhaustive extracted of <i>C. padoides</i> leaves.	
Table 3-3: Total number of UV and vanillin spray reagent active compounds resulting from	n direct extraction
with acetone of <i>C. imberbe</i> and <i>C. padoides</i>	46
Table 3-4: Total number of UV and vanillin sulphuric acid spray reagent active compounds serial exhaustive extraction extracts of <i>C. imberbe</i> and <i>C. padoides</i> in EMV	/ solvent system
Table 4-1: Bioautography (TLC in EMW) and Minimim Inhibitrory Concentration (MIC) results screening of <i>C. imberbe</i> , <i>C. padoides</i> , <i>C. celastroides</i> ssp. <i>orientale</i> and <i>C. celastroides</i> acetone extracts against <i>S. aureus</i> (SA), <i>E. faecalis</i> (EF), <i>E. aeruginosa</i> (PA)	celastroides ssp
Table 4-2: Bioautography, MIC and total activity values of serial exhaustive extracts ( <i>S faecalis</i> (EF), <i>E. coli</i> (EC) and <i>P. aeruginosa</i> (PA)	
Table 5-1: Gradients of solvent used in VLC for the separation of the dichlorome <i>C.imberbe</i>	
Table 5-2: Quantity (g) and percentage of initial mass (10 g) of <i>C. imberbe</i> and <i>C. padoio</i> extracted by different solvents in a solvent-solvent fractionation process	
Table 5-3: Quantity obtained, colour and complexity of fractions resulting from a VLC pro- extract of <i>C. imberbe</i>	

Table 5-4: Quant	tity of fraction	ons derive	d through gravit	y gradient	column	chromat	ography	of the ch	nlorof	orm
	obtained		solvent-solven							
Table 5-5: Number	er of antibac	terial com	oounds and MIC	values of fr	actions r	resulting	from the	solvent-	solvei	nt
fractiona	ation process	s of the DC	CM fraction of C.	<i>imberbe</i> an	ıd <i>C.pad</i>	oides				80
Table 5-6: Quant	tity obtained	I, no of co	ompounds, and	MIC of VL	C fraction	ns of Do	CM extra	act of C.	imbe	<i>rb</i> e
										81
Table 5-7: Derepl	lication, R <sub>f</sub> v	alues of is	olated compound	ds						.83
Table 6-1: <sup>13</sup> C (30	00 MHz) and	I <sup>13</sup> C (75.4	MHz) NMR data	of 1-4 (CD	₃OD)				<i>'</i>	101
Table 6-2: <sup>1</sup> H (30	00 MHz) and	<sup>13</sup> C (75.4	MHz) NMR data	of 6 (CD <sub>3</sub> O	D)					104
Table 6-3. HMBC	correlations	s for methy	d groups in comp	ound 5					<i>'</i>	105
Table 6-4: 13C (75	5.4 MHz) an	d <sup>1</sup> H (300 I	MHz) NMR data	of 7 band 8	in CD₃C	D			1	117
Table 6-5: <sup>1</sup> H (30	00 MHz) and	<sup>13</sup> C (75.4	MHz) NMR data	of 6(CD <sub>3</sub> )					1	18
Table 7-1: Bacte	ria and Funເ	gi tested fo	r effficacy in the	Agar diffus	ion assa	y				122
Table 7-2: MIC of	f compounds	s isolated f	from C. imberbe	and <i>C. pad</i>	loides [S	. aureus	(SA), <i>E.</i>	faecalis (	(EF),	and
E. coli (E	EC), P. aeru	<i>ginosa</i> (P <i>F</i>	۸)]						··········	125
Table 7-3: Zone of	of inhibition of	of compou	nds isolated fron	n <i>C. imberb</i>	e and C.	padoide	es agains	st several	bacte	eria
and fung	gal organism	ı <b>s</b> ( <i>B. subt</i>	ilis ATTC 6633 (	MET) NA (I	B1), <i>B. s</i>	ubtilis A	TTC 663	3 (IMET)	NS (I	B2)
S. aure	us (IMET 1	0760) SG	511 (B3), <i>E. c</i>	oli SG 458	B (B4), /	P. aerug	<i>ginosa</i> K	799/61	(B9),	M
o .		•	056) (M2), <i>N</i>				` ,			
Table 7-4: MIC (										
Table 7-5: Anti	,				· ·					
Table 7-6: Anti-pr										