
1

Introduction to Parasitic Plants

Abstract

Parasitic plants are generally little known by agriculturalists even though under some conditions they may be the most important factor in crop losses. Globally, they have their greatest impact on food crops of smallholder farming systems in Africa. Despite this, they are often treated simply as weeds, overlooking the fact that they do not just harm the crop indirectly through competition, but also directly through parasitism. Parasitic weeds connect to crop plants through a specialized structure, the haustorium. In fact, the presence of a haustorium is what defines a parasitic plant. Parasitism has arisen in 12 clades of angiosperms, yielding plants with a diversity of habits including herbs, vines, shrubs and even trees. Likewise, there is a range of parasite–host interactions. Some parasites will only germinate with a stimulant produced by the host. Some are specific in host selection, some are promiscuous with many different hosts, and some are not quite generalists but are not host specific. We include all known African parasites that attack crops, with emphasis on mistletoes, witchweeds, dodders and broomrapes, including their taxonomy, hosts, distribution and control measures.

1.1 Parasitic Plants as Weed Problems

Parasitism has been reported in 28 plant families, comprising nearly 4500 species, all exclusively dicotyledons (Heide-Jørgensen, 2013; Nickrent, 2020). When these plants parasitize other plants, either out of necessity or to increase their reproductive output (Shen *et al.*, 2006), the host plants can be severely damaged. When hosts are agricultural crops, parasitic plants can become important weed problems. A broad range of African crops suffer from parasitic weeds. Affected crops include staple food grains (e.g. maize, rice, sorghum, millet) and legumes (e.g. cowpea, faba beans, lentils), a diversity of vegetables (e.g. carrots, tomatoes, leek), oil crops (e.g. sunflower, linseed), fibre crops (e.g. flax, hemp), forage crops (e.g. lucerne, clover), many fruit-tree species (e.g. mango, guava, citrus) and plantation cash crops (e.g. cacao, coffee, tea, rubber).

Parasitic plants can lead to severe yield losses, making them an important constraint to food security in many areas (Fig. 1.1). While quantitative information on yield losses from parasitic weeds is lacking for many parasite–host species combinations, available data emphasize just how serious these pathogens are. An assessment by Rodenburg *et al.* (2016a) showed that when *Striga asiatica* is not controlled, mean yield losses of upland rice are around 73%. For maize, the same parasite causes yield losses of 80% or higher when uncontrolled (Ransom *et al.*, 1990; Rusinamhodzi *et al.*, 2012). *Striga hermonthica* can cause yield losses of up to 84% (mean: 37%) in sorghum (Rodenburg *et al.*, 2005) and up to 81% in maize (mean: 68%; Kim *et al.*, 2002), depending on variety, infestation level and environmental conditions. *Rhizophora fistulosa* causes yield losses of rice ranging from 24% to 73% (mean: 50%), again depending on the variety and infestation level (Rodenburg *et al.*, 2016b). Field dodder, *Cuscuta campestris*, reduces yields of sesame by 67%, soybean by 48%, pigeon pea by 25% and groundnut by 18% (Mishra *et al.*, 2007). *Alectra vogelii* inflicted yield losses in susceptible cowpea varieties that were reported to range from 30% to 66% (mean: 51%; Alonge *et al.*, 2001) whereas *Striga gesnerioides* inflicted yield losses that ranged from 79% to 86% (mean: 81%; Alonge *et al.*, 2005), but for both parasite species, these losses were reduced in some of the resistant and tolerant cowpea genotypes. Yield losses caused by broomrapes (*Orobancha* spp. and *Phelipanche* spp.) in faba bean, chickpea, tomato, potato and sunflower range from 5% to 100% (Abang *et al.*, 2007).

No quantitative data exist on damage inflicted by mistletoe but assessment is based on field observations and farmer perceptions. Loranthaceae parasitism causes important shea tree yield reductions in Burkina Faso (Boussim *et al.*, 2004). Damage to these and other economically important crops is generally increased by low soil fertility and drought stress, conditions facing many African smallholders.

The above yield-loss estimates are field- or crop-scale measurements. The extent of the parasitic weed problem in Africa cannot be truly assessed without quantitative information on the spread of the different parasite species across croplands and their economic impact at a national and regional scale. The data on parasitic weed distribution and economic impact in Africa are scarce, however, and are mainly associated with those parasites that impact the region's cereal production. Maize cropland infested by *Striga* spp. (chiefly *Striga hermonthica* and *S. asiatica*) in sub-Saharan Africa is estimated at 2.3 million ha and the concomitant annual economic losses are estimated at US\$383 million (Woomer *et al.*, 2008). The area of rainfed rice infested by parasitic weeds (*Striga hermonthica*, *S. asiatica*, *S. aspera* or *Rhizophora fistulosa*) is estimated at 1.34 million ha (about 19% of the total area under rainfed rice) resulting in a total estimated annual economic impact of at least US\$111 million (Rodenburg *et al.*, 2016a). The total annual loss caused by *S. hermonthica*, one of the main parasitic weeds in cereals in Africa, is roughly estimated to be more than US\$1 billion (Parker, 2009). For Africa, no quantitative economic impact data are available on any of the stem parasites described in this book.



Fig. 1.1. Farmers in parasitic-weed-infested field crops in Africa. (A) Rice field infested by *Striga hermonthica* (purple-flowered plants) in Côte d'Ivoire. (B) Rice field infested by *Rhamphicarpa fistulosa* (reddish plants among the rice) in Uganda.

1.2 What is a Parasitic Plant?

Although parasitic plants are often thought of as weeds, they are part of a guild of highly unique plants, the parasitic angiosperms. An understanding of their biology is essential for effective control and management. Parasitic plants are amazingly specialized, with remarkable adaptations for their heterotrophic existence. Their habits are diverse, including herbaceous plants, vines, shrubs and trees. Some appear innocuous, with no external evidence of their parasitic nature. Others lack chlorophyll or even leaves and stems, existing only within the bodies of other plants until they flower. Parasitic plants' reproductive strategies also vary widely, from the tiny (1 mm) flowers of some mistletoes to the metre-wide flowers of *Rafflesia* species – the largest flower in the world. Unique among African parasitic plants is the rainforest tree *Okoubaka aubrevillei* (Santalaceae), a rare but widely distributed tree in Western and Central Africa, much sought after for its purported medicinal value. It is the largest parasitic plant in the world and little studied. Veenendaal *et al.* (1996) present the only data from experimental work on host selection and host damage. In their study, they found that *O. aubrevillei* caused morbidity and death in seedlings of *Pericopsis elata*, a leguminous rainforest tree. The authors suggest that *O. aubrevillei* favours such nitrogen-fixing trees and that the role of parasitism is to reduce competition at the seedling stage.

What this diverse coterie of plants share is a haustorium. Simply put, if a haustorium is present, the plant is a parasite. It is the defining feature of this group of organisms. The haustorium is the morphological and physiological bridge between host and parasite. This structure is the conduit for water and dissolved materials, such as nutrients and metabolites, but also proteins and pathogens (Yoshida *et al.*, 2016) as well as genetic material transported from the host into the parasite or from the parasite into the host. Non-parasitic weeds compete with crop plants for water and nutrients in the soil, whereas parasitic weeds obtain these resources directly from host plants. Farmers are sometimes surprised to learn that some of the weeds in their crops, in particular the ones with green leaves such as witchweeds, are also parasites. Knowing the parasitic behaviour is, however, essential to understanding control measures.

1.3 Categories of Parasitic Weeds

There are roughly four different categories of parasitic plants (Table 1.1). Parasitic plants can be distinguished by the presence or absence of chlorophyll. Those that produce chlorophyll (and therefore have some photosynthetic activity) are termed hemiparasites (also known as semiparasites), and this category comprises about 90% of all parasitic plant species (Heide-Jørgensen, 2013). Those that lack chlorophyll (and therefore are not green and are totally dependent upon their host for nutrition and water) are termed holoparasites. Another distinction among parasites is with germination. Obligate parasites require the presence of a host to germinate and initiate a haustorium. Facultative parasites, on the other hand, can germinate without a host (see Kabiri *et al.*, 2016).

Table 1.1. Parasitic plant species reported to be weed problems in African agriculture.

Parasitism and common name	Family	Genus	Species	Main crop hosts	Chapter ^a
Stem parasites					
Obligate hemiparasites					
Mistletoe	Loranthaceae	<i>Tapinanthus</i>	<i>T. bangwensis</i> <i>T. belvisii</i>	Guava, other tree crops Tree crops	2 2
		<i>Erianthemum</i>	<i>E. dregei</i>	Tree crops	2
		<i>Phragmanthera</i>	<i>P. capitata</i> <i>P. incana</i>	Tree crops	2 2
	Viscaceae	<i>Viscum</i>	<i>V. cruciatum</i> <i>V. anceps</i> <i>V. engleri</i> <i>V. rotundifolium</i>	Tree crops	2 2 2 2
Love vine	Lauraceae	<i>Cassytha</i>	<i>C. filiformis</i>	Mango, cashew, other tree crops	3
Dodder	Convolvulaceae	<i>Cuscuta</i>	<i>C. campestris</i> <i>C. epilinum</i> <i>C. suaveolens</i> <i>C. hyalina</i> <i>C. monogyna</i> <i>C. epithymum</i> <i>C. pedicellata</i> <i>C. planiflora</i> <i>C. australis</i> <i>C. chinensis</i> <i>C. kilimanjari</i>	Vegetables, tree crops, forage crops Flax Tree crops Forage crops, lucerne Vetch, lentil, arugula Lucerne Lucerne Various Cassava, ornamental shrubs	4 4 4 4 4 4 4 4 4 4 4
Field dodder					
Fringed dodder					
Eastern dodder					
Lucerne dodder					
Australian dodder					
Chinese dodder					

Continued

Table 1.1. Continued.

Parasitism and common name	Family	Genus	Species	Main crop hosts	Chapter ^a			
Root parasites								
Facultative hemiparasites								
Rice vampire weed	Orobanchaceae	Rhamphicarpa	R. fistulosa	Rice, maize	5			
			<i>R. brevipedicellata</i>		5			
			<i>R. capillacea</i>		5			
			<i>R. elongata</i>		5			
			<i>R. veronicaefolia</i>		5			
Hairy buchnera	Orobanchaceae	<i>Buchnera</i>	<i>B. hispida</i>	Cereals	6			
NA	Orobanchaceae							
		<i>Micrargeria</i>	<i>M. filiformis</i>	Rice	11			
		<i>Sopubia</i>	<i>S. parviflora</i>	Rice	11			
Thesium	Santalaceae/ Thesiaceae	<i>Thesium</i>	<i>T. humile</i>		11			
			<i>T. resedoides</i>		11			
Obligate hemiparasites								
Witchweed	Orobanchaceae	Striga	S. asiatica	Sorghum, millet, maize, rice, sugarcane	7			
Red witchweed								
Witchweed						<i>S. aspera</i>	Cereals, fonio, sugarcane	7
Purple witchweed						S. hermonthica	Sorghum, millet, maize, rice, sugarcane	7
Cowpea witchweed						S. gesnerioides	Cowpea, sweet potato, tobacco	7
Giant maize witchweed						<i>S. forbesii</i>	Cereals, sugarcane	7
NA						<i>S. passargei</i>	Cereals	7
NA		<i>S. brachycalyx</i>	Cereals	7				
Alectra	Orobanchaceae							

		Alectra	A. vogelii	Cowpea, groundnut	8
			<i>A. picta</i>	Pulses	8
			<i>A. sessiliflora</i>	Pulses	8
			<i>A. orobanchoides</i>	Sunflower, tobacco	8
Obligate holoparasites					
Broomrape	Orobanchaceae				
Bean broomrape		<i>Orobanche</i>	<i>O. crenata</i>	Faba bean, bean, chickpea	9
Sunflower broomrape			<i>O. cumana</i> syn.	Sunflower, tobacco, tomato,	9
			<i>cernua</i>	aubergine	
Small broomrape			<i>O. minor</i>	Tobacco, lettuce, forage legumes	9
Stinking broomrape			<i>O. foetida</i>	Pulses	9
Egyptian broomrape		<i>Phelipanche</i>	<i>P. aegyptiaca</i>	Potato, tomato, melons	9
Branched broomrape			<i>P. ramosa</i>	Potato, tomato, aubergine, tobacco,	9
				cole crops	
Thonningia	Balanophoraceae	<i>Thonningia</i>	<i>T. sanguinea</i>	Tree crops including rubber, coffee and cacao	10

Species in bold are the most economically important.

^aChapter in this volume where the species is discussed.

NA = no widely accepted common name available.

Intuitively, it seems that the most serious parasitic weeds would be holoparasites. And indeed, species of *Orobancha* and *Phelipanche* are well-known pathogens of a variety of crops. But in Africa, the most serious parasitic weeds are the witchweeds, which are obligate hemiparasites in the genus *Striga*. A further broad distinction can be made between categories of parasitic weeds in terms of where they parasitize their hosts. Around 40% of parasitic plants attack stems, whereas others are restricted to roots. These are simply referred to respectively as stem parasites and root parasites.

1.4 Parasitic Plant Research

The modern science of parasitic plants was launched in 1969 by the publication of Job Kuijt's magisterial biology of parasitic plants (Kuijt, 1969). This drew attention to a group of plants known chiefly for their bizarre morphology. A decade earlier, in-depth studies on physiology, biochemistry and control were stimulated by the discovery of *Striga asiatica* (red witchweed) in North and South Carolina (USA) in the 1950s. The parasite quickly developed as a serious pathogen of maize in these states, prompting extensive work on the biology, control and containment of this species. As a result, after many years of work, the elegant, complex germination biology of witchweed and other parasites has been elucidated and parasitic plant research expanded worldwide, leading to a surge in publications on parasitic plants.

Following Kuijt's treatment, a series of books on parasitic plants has appeared, for example Parker and Riches (1993), Press and Graves (1995), Heide-Jørgensen (2008) and Joel *et al.* (2013). These volumes deal with parasitic angiosperms as a whole. Less exhaustive discussions of parasitic plants are summarized in Těšitel (2016), Nickrent and Musselman (2017), and Teixeira-Costa and Davis (2021). Reviews of groups of parasites we cover in this book can be found in their respective chapters.

As the number of publications suggests, an appraisal and review of research would be a large undertaking, beyond the scope of this book. As examples, two highlights stand out: first, phylogenetic studies, well reviewed in Nickrent (2020) documenting the evolution of parasitism in 12 clades of angiosperms; and second, the germination biology of parasites, especially root parasites. This has resulted in the discovery of a new group of plant hormones, the strigolactones. These growth regulators are now known to be widespread in angiosperms. A helpful review of strigolactones is provided by Xie *et al.* (2010).

The heightened level of research in parasitic plants is now a worldwide phenomenon. In 1957, in response to the discovery of witchweed in the USA, an exhaustive review of the world literature on witchweed was published as a detailed annotated bibliography (McGrath *et al.*, 1957). It had 298 references, including non-peer-reviewed entries. A November 2022 Web of Science search (all peer reviewed) for *Striga* yielded 1801 strikes. Similarly, an extensive review of *Cuscuta* in 1994 (Dawson *et al.*, 1994) had 303 references, the Web of Science search for *Cuscuta* gave 1271, and for *Orobancha* (including *Phelipanche*) (broomrapes) about 1585. Of course, not all the references

concern agriculture or even biology. Studies on these plants have expanded beyond agronomic interest to phylogenetic research, physiology, herbal medicines, ecology and more.

Despite the thousands of studies by scientists around the world, smallholder farmers in Africa have profited little by the effort and expense put into understanding parasitic weeds. Control, either by reducing infestations or by reducing the impact on the host, is seldom realized by the farmer whose management of the parasites affects daily existence. It has been previously observed by Schut *et al.* (2015a) that research on parasitic weeds in Africa has mainly focused on understanding the biology, ecology and distribution of the parasites, and on the development and testing of strategies for managing them, with some efforts on understanding the socio-cultural dimension (e.g. Vissoh *et al.*, 2008; N'cho *et al.*, 2014) and economic impact of parasitic weeds (e.g. N'cho *et al.*, 2017, 2019). The institutional and political dimensions of parasitic weeds and the innovations to address them have not received the same structural attention. While farmers frequently participate in parasitic weed research (e.g. Schulz *et al.*, 2003; Emechebe *et al.*, 2004; Abang *et al.*, 2007; Tippe *et al.*, 2017), the private sector, civil society organizations and government representatives are less often involved (Schut *et al.*, 2015b). For research on parasitic weeds to benefit smallholder farmers, involving a broader range of stakeholders and considering broader dimensions than just the crop or farm is deemed necessary (Rodenburg *et al.*, 2015).

1.5 Parasitic Weeds in African Agricultural Systems

Parasitic weed infestation, in particular by species of the Orobanchaceae, constitutes one of the most important and complex agricultural production constraints in Africa (e.g. Vurro *et al.*, 2010; Waddington *et al.*, 2010). The problem is important because staple crops such as maize, rice, sorghum and millet are important hosts of a number of the parasitic weed species (e.g. *Striga hermonthica*, *S. asiatica*, *Rhamphicarpa fistulosa*) and because these species are widely distributed (e.g. Rodenburg *et al.*, 2016b). Hence, the parasitic weed problem greatly affects food security in the region.

The problem is complex because of the ingenious biology of plant parasitism (see Shen *et al.*, 2006; Spallek *et al.*, 2013; Těšitel, 2016). Many weedy species of parasitic plants have a wide host range, and their germination and reproductive biology render them highly successful in annually cropped environments. The problem is also difficult because most of the affected crops in Africa are predominantly grown by smallholder farmers. Although smallholder farming systems in Africa are highly diverse in their resources, environments, challenges and opportunities (e.g. Tittonell *et al.*, 2010), the majority of farmers struggle with adverse environmental conditions and limited access to productive agricultural land, production resources, information and services. These conditions render the control of parasitic weeds an even more difficult task.

Parasitic weed infection and damage is often associated with and aggravated by adverse biophysical conditions such as poor soil fertility and drought. The

weeds present technological challenges because the number of feasible, effective and affordable control measures is limited (e.g. Tippe *et al.*, 2017; Silberg *et al.*, 2020) or farmers are unaware of them. The affordability, accessibility and awareness of control strategies are a direct function of the socio-cultural, economic, institutional and even political dimensions shaping this problem; agricultural extension services in rural Africa are often poorly staffed, poorly equipped and ill-informed on parasitic weed problems and ways to address them, and communications between farmers and extension and crop protection services are often suboptimal (Schut *et al.*, 2015b). Therefore, addressing the problem of parasitic weeds in Africa, by technological and organizational control strategies, requires not only a thorough understanding of the biology and ecology of the important species but also a better understanding of the social, economic and institutional environments where these weeds are problems. Such research and development endeavours need to involve a range of stakeholders, including social and natural science researchers, farmers, extension services, and public and private crop health services. The control strategies arising from such a transdisciplinary research approach should match the resource availability and farming practices of the farmers who need to implement them and should be effectively communicated to them and be locally available at an affordable price or input level.

The present work deals with parasitic plants that are current or potential agricultural pests (Table 1.1). Although it is beyond the scope of this book to note them all, parasitic species that are not currently a problem in Africa possess – at least theoretically – the ability to become weedy and cause crop damage in the future. There are examples of indigenous parasitic plants becoming pathogens in agriculture and forestry (e.g. *Thonningia sanguinea* on rubber, coffee and other crops in Western Africa; Imarhiagbe and Aigbokhan, 2019). Knowledge on biology and control of those species representing current parasitic weed problems in Africa, as well as on the socio-economic and institutional environments of farming systems where these problems are embedded, could prepare us for future outbreaks. The hope of the authors is that this contribution will increase the awareness of these plants as parasitic pathogens – especially those that are currently lesser known – ultimately to aid the smallholder farmer in Africa.

References

- Abang, M.M., Bayaa, B., Abu-Irmaileh, B. and Yahyau, A. (2007) A participatory farming system approach for sustainable broomrape (*Orobancha* spp.) management in the Near East and North Africa. *Crop Protection* 26, 1723–1732.
- Alonge, S.O., Lagoke, S.T.O. and Ajakaiye, C.O. (2001) Cowpea reactions to *Alectra vogelii* II: effect on yield and nutrient composition. *Crop Protection* 20, 291–296.
- Alonge, S.O., Lagoke, S.T.O. and Ajakaiye, C.O. (2005) Cowpea reactions to *Striga gesnerioides* II. Effect on grain yield and nutrient composition. *Crop Protection* 24, 575–580.
- Boussim, I.J., Guinko, S., Tuquet, C. and Sallé, G. (2004) Mistletoes of the agroforestry parklands of Burkina Faso. *Agroforestry Systems* 60, 39–49.
- Dawson, J.H., Musselman, L.J., Wolswinkel, P. and Dörr, I. (1994) Biology and control of *Cuscuta*. *Reviews of Weed Science* 6, 265–317.

- Emechebe, A.M., Ellis Jones, J., Schulz, S., Chikoye, D., Douthwaite, B. *et al.* (2004) Farmers' perception of the *Striga* problem and its control in Northern Nigeria. *Experimental Agriculture* 40, 215–232.
- Heide-Jørgensen, H.S. (2008) *Parasitic Flowering Plants*. Brill, Leiden, The Netherlands.
- Heide-Jørgensen, H.S. (2013) Introduction: the parasitic syndrome in higher plants. In: Joel D.M., Gressel J. and Musselman, L.J. (eds) *Parasitic Orobanchaceae*. Springer, Berlin, pp. 1–18.
- Imarhiagbe, O. and Aigbokhan, E.I. (2019) Studies on *Thonningia sanguinea* Vahl (Balanophoraceae) in southern Nigeria. Range and host preference. *International Journal of Conservation Science* 10, 721–732.
- Joel, D.M., Gressel, J. and Musselman, L.J. (eds) (2013) *Parasitic Orobanchaceae: Parasitic Mechanisms and Control Strategies*. Springer, Berlin.
- Kabiri, S., Van Ast, A., Rodenburg, J. and Bastiaans, L. (2016) Host influence on germination and reproduction of the facultative hemi-parasitic weed *Rhamphicarpa fistulosa*. *Annals of Applied Biology* 169, 144–154.
- Kim, S.K., Adetimirin, V.O., The, C. and Dossou, R. (2002) Yield losses in maize due to *Striga hermonthica* in West and Central Africa. *International Journal of Pest Management* 48, 211–217.
- Kuijt, J. (1969) *The Biology of Parasitic Flowering Plants*. University of California Press, Berkeley, California.
- McGrath, H., Shaw, W.C., Jansen, L.L., Lipscomb, B.R., Miller, P.R. *et al.* (1957) *Witchweed (Striga asiatica) – A New Parasitic Plant in the United States*. US Department of Agriculture, Special Publication 10, Washington, DC.
- Mishra, J.S., Moorthy, B.T.S., Bhan, M. and Yaduraju, N.T. (2007) Relative tolerance of rainy season crops to field dodder (*Cuscuta campestris*) and its management in niger (*Guizotia abyssinica*). *Crop Protection* 26, 625–629.
- N'Cho, S.A., Mourits, M., Rodenburg, J., Demont, M. and Lansink, A.O. (2014) Determinants of parasitic weed infestation in rainfed lowland rice in Benin. *Agricultural Systems* 130, 105–115.
- N'Cho, S.A., Mourits, M., Demont, M., Adegbola, P.Y. and Lansink, A.O. (2017) Impact of infestation by parasitic weeds on rice farmers' productivity and technical efficiency in sub-Saharan Africa. *African Journal of Agricultural and Resource Economics* 12, 35–50.
- N'Cho, S.A., Mourits, M., Rodenburg, J. and Lansink, A.O. (2019) Inefficiency of manual weeding in rainfed rice systems affected by parasitic weeds. *Agricultural Economics* 50, 151–163.
- Nickrent, D.L. (2020) Parasitic angiosperms: how often and how many? *Taxon* 69, 5–27.
- Nickrent, D.L. and Musselman, L.J. (2017) Parasitic plants. In: Ownley, B.H. and Trigiano, R.N. (eds) *Plant Pathology: Concepts and Laboratory Exercises, 3rd edn*. CRC Press, Boca Raton, Florida, pp. 277–288.
- Parker, C. (2009) Observations on the current status of *Orobanche* and *Striga* problems worldwide. *Pest Management Science* 65, 453–459.
- Parker, C. and Riches, C.R. (1993) *Parasitic Weeds of the World*. CAB International, Wallingford, UK.
- Press, M.C. and Graves, J.D. (eds) (1995) *Parasitic Plants*. Chapman and Hall, London.
- Ransom, J.K., Eplee, R.E. and Langston, M.A. (1990) Genetic variability for resistance to *Striga asiatica* in maize. *Cereal Research Communications* 18, 329–334.
- Rodenburg, J., Bastiaans, L., Weltzien, E. and Hess, D.E. (2005) How can field selection for *Striga* resistance and tolerance in sorghum be improved? *Field Crops Research* 93, 34–50.
- Rodenburg, J., Schut, M., Demont, M., Klerkx, L., Gbehounou, G. *et al.* (2015) Systems approaches to innovation in pest management: reflections and lessons learned from an integrated research program on parasitic weeds in rice. *International Journal of Pest Management* 61, 329–339.
- Rodenburg, J., Cissoko, M., Dieng, I., Kayeke, J. and Bastiaans, L. (2016a) Rice yields under *Rhamphicarpa fistulosa*-infested field conditions, and variety selection criteria for resistance and tolerance. *Field Crops Research* 194, 21–30.

- Rodenburg, J., Demont, M., Zwart, S.J. and Bastiaans, L. (2016b) Parasitic weed incidence and related economic losses in rice in Africa. *Agriculture, Ecosystems & Environment* 235, 306–317.
- Rusinamhodzi, L., Corbeels, M., Nyamangara, J. and Giller, K.E. (2012) Maize–grain legume intercropping is an attractive option for ecological intensification that reduces climatic risk for smallholder farmers in central Mozambique. *Field Crops Research* 136, 12–22.
- Schulz, S., Hussaini, M.A., Kling, J.G., Berner, D.K. and Ikie, F.O. (2003) Evaluation of integrated *Striga hermonthica* control technologies under farmer management. *Experimental Agriculture* 39, 99–108.
- Schut, M., Rodenburg, J., Klerkx, L., Kayeke, J., Van Ast, A. *et al.* (2015a) RAAIS: Rapid Appraisal of Agricultural Innovation Systems (Part II). Integrated analysis of parasitic weed problems in rice in Tanzania. *Agricultural Systems* 132, 12–24.
- Schut, M., Rodenburg, J., Klerkx, L., Hinnou, L.C., Kayeke, J. *et al.* (2015b) Participatory appraisal of institutional and political constraints and opportunities for innovation to address parasitic weeds in rice. *Crop Protection* 74, 158–170.
- Shen, H., Ye, W., Hong, L., Huang, H., Wang, Z. *et al.* (2006) Progress in parasitic plant biology: host selection and nutrient transfer. *Plant Biology* 8, 175–185.
- Silberg, T.R., Richardson, R.B. and Lopez, M.C. (2020) Maize farmer preferences for intercropping systems to reduce *Striga* in Malawi. *Food Security* 12, 269–283.
- Spallek, T., Mutuku, M. and Shirasu, K. (2013) The genus *Striga*: a witch profile. *Molecular Plant Pathology* 14, 861–869.
- Těšitel, J. (2016) Functional biology of parasitic plants: a review. *Plant Ecology and Evolution* 149, 5–20.
- Texeira-Costa, L. and Davis, C.C. (2021) Life history, diversity, and distribution of parasitic flowering plants. *Plant Physiology* 187, 32–51.
- Tippe, D.E., Rodenburg, J., Schut, M., Van Ast, A., Kayeke, J. *et al.* (2017) Farmers' knowledge, use and preferences of parasitic weed management strategies in rain-fed rice production systems. *Crop Protection* 99, 93–107.
- Tittonell, P., Muriuki, A., Shepherd, K.D., Mugendi, D., Kaizzi, K.C. *et al.* (2010) The diversity of rural livelihoods and their influence on soil fertility in agricultural systems of East Africa – a typology of smallholder farms. *Agricultural Systems* 103, 83–97.
- Veenendaal, E.M., Aberese, I.K., Walsh, M.F. and Swaine, M.D. (1996) Root parasitism in a West African rainforest tree *Okoubaka aubrevillei* (Santalaceae). *New Phytologist* 134, 487–493.
- Vissoh, P.V., Gbèhounou, G., Ahanchede, A., Roling, N.G. and Kuyper, T.W. (2008) Evaluation of integrated crop management strategies employed to cope with *Striga* infestation in permanent land use systems in southern Benin. *International Journal of Pest Management* 54, 197–206.
- Vurro, M., Bonciani, B. and Vannacci, G. (2010) Emerging infectious diseases of crop plants in developing countries: impact on agriculture and socio-economic consequences. *Food Security* 2, 113–132.
- Waddington, S.R., Li, X.Y., Dixon, J., Hyman, G. and de Vicente, M.C. (2010) Getting the focus right: production constraints for six major food crops in Asian and African farming systems. *Food Security* 2, 27–48.
- Woomer, P.L., Bokanga, M. and Odhiambo, G.D. (2008) *Striga* management and the African farmer. *Outlook on Agriculture* 37, 277–282.
- Xie, S., Yoneyama, K. and Yoneyama, K. (2010) The strigolactone story. *Annual Review of Phytopathology* 48, 93–117.
- Yoshida, S., Cui, S.K., Ichihashi, Y. and Shirasu, K. (2016) The haustorium, a specialized invasive organ in parasitic plants. *Annual Review of Plant Biology* 67, 643–667.